WE CLAIM:

1. A method of *in vitro* cultivation of *Polygonatum* comprising

contacting a *Polygonatum* seed with a first medium comprising a MS basal culture medium and from about 10 mg/L to about 100 mg/L gibberellic acid (GA_3);

upon emergence of a hypocotyl, contacting this primary explant with a second medium comprising a MS basal culture medium, from about 3 mg/L to about 6 mg/L of 6-benzyl-aminopurine, and from 0.5 mg/L to about 1.0 mg/L naphthalene acetic acid ; and

upon emergence of a first foliage leaf, contacting this secondary explant with a third medium comprising a MS basal culture medium, about 2.0 mg/L 6-benzyl-aminopurine, about 1.0 mg/L naphthalene acetic acid, and from about 5 mg/L to about 20 mg/L gibberellic acid (GA_3).

2. The method of claim 1, wherein the *Polygonatum* is selected from the group consisting of *Polygonatum cirrhifolium*, *Polygonatum oppositifolium*, and *Polygonatum verticillatum* L.

3. The method of claim 2, wherein the *Polygonatum cirrhifolium*, is *Polygonatum cirrhifolium* Royle.

4. The method of claim 2, wherein the *Polygonatum oppositifolium*, is *Polygonatum oppositifolium* Royle.

5. The method of claim 1, wherein the concentration of GA_3 in the first media is from about 50 mg/L to about 100 mg/L.

6. The method of claim 1, wherein the concentration of NAA in the second media is about 1.0 mg/L.

7. The method of claim 1, wherein the concentration of GA_3 in the third media is from about 15 mg/L to about 20 mg/L GA_3.

8. The method of claim 1, wherein the MS basal culture medium of the first, second, and/or third medium is MS basal culture medium I.

9. The method of claim 1, wherein the MS basal culture medium of the first, second, and/or third media is MS basal culture medium II.

10. The method of claim 1, wherein the first, second, and/or third media further comprise agar.

11. The method of claim 1, wherein the MS basal culture medium of the first, second, and/or third medium has a pH of about 5.8.

12. The method of claim 1, wherein the temperature is from about 20° C to about 24° C.

13. The method of claim 1, wherein the relative humidity is about 50% to about 60%.

14. The method of claim 1, wherein said *Polygonatum* is contacted with said first medium in the substantial absence of light.

15. The method of claim 1, wherein said *Polygonatum* is contacted with said second and/or third medium under illumination of about 2000 to about 3500 lux.

16. The method of claim 1, wherein the primary explant is contacted with the second medium less than 60 days after said seeds are contacted with the first medium.

17. A method of *in vitro* cultivation of *Polygonatum cirrhifolium* Royle comprising the steps of contacting a *Polygonatum cirrhifolium* Royle seed with a first medium comprising a MS basal culture medium and from about 10 mg/L to about 100 mg/L gibberellic acid (GA_3);

upon emergence of a hypocotyl, contacting this primary explant with a second medium comprising a MS basal culture medium, from about 3 mg/L to about 6 mg/L of 6-

benzyl-aminopurine, and from 0.5 mg/L to about 1.0 mg/L naphthalene acetic acid ; and

upon emergence of a first foliage leaf, contacting this secondary explant with a third medium comprising a MS basal culture medium, about 2.0 mg/L 6-benzyl-aminopurine, about 1.0 mg/L naphthalene acetic acid, and from about 5 mg/L to about 20 mg/L gibberellic acid (GA_3).

wherein the temperature is from about 20° C to about 24° C, and the relative humidity is about 50% to about 60%, the hypocotyl-bearing seed is contacted with the second medium less than 60 days after said seeds are contacted with the first medium, and the seedling is contacted with the third medium less than 90 days after said seeds are contacted with the first medium.

18. A method of germinating 65-100% of *Polygonatum* seeds in less than 60 days comprising contacting a plurality of *Polygonatum* seeds with a first medium comprising a MS basal culture medium and from about 10 mg/L to about 100 mg/L gibberellic acid (GA_3).

19. The method of claim 12, wherein the germination rate is about 98% and the concentration of gibberillic acid (GA_3) is about 50 mg/L.

20. A method of germinating *Polygonatum* seeds comprising contacting a plurality of *Polygonatum* seeds with a first medium comprising a MS basal culture medium and from about 50 mg/L gibberellic acid (GA_3) wherein 65-100% of said *Polygonatum* seeds germinate in less than 60 days.

21. The method of claim 14, wherein the germination rate is about 98% and the concentration of gibberillic acid (GA_3) is about 50 mg/L.

22. A method of inducing synchronous release of epicotyl dormancy, coleoptile dormancy, and radicle dormancy in *Polygonatum* comprising contacting a hypocotyl-bearing *Polygonatum* seed with a medium comprising a MS basal culture medium, from about 3 mg/L to about 6 mg/L of 6-benzyl-aminopurine, and from 0.5 mg/L to about 1.0 mg/L naphthalene acetic acid.

23. A kit comprising a package insert and

a first medium comprising MS basal culture medium and from about 10 mg/L to about 100 mg/L Gibberellic acid (GA_3);

a second medium comprising from about 3 mg/L to about 6 mg/L of 6- benzyl-aminopurine (BAP), and from 0.5 mg/L to about 1.0 mg/L naphthalene acetic acid ; and

a third medium comprising about 2.0 mg/L BAP, about 1.0 mg/L NAA, and from about 5 mg/L to about 20 mg/L GA_3.

24. A kit comprising a package insert and

a germination medium comprising MS basal culture medium and from about 10 mg/L to about 100 mg/L Gibberellic acid (GA_3);

a leafing medium comprising from about 3 mg/L to about 6 mg/L of 6- benzyl-aminopurine (BAP), and from 0.5 mg/L to about 1.0 mg/L naphthalene acetic acid ; and

a budding medium comprising about 2.0 mg/L BAP, about 1.0 mg/L NAA, and from about 5 mg/L to about 20 mg/L GA_3.

25. Culture media compositions useful for initiating extraordinarily fast and synchronized *in vitro* induction of germination and release of epicotyl dormancy in *Polygonatum cirrhifolium*

Royle, an endangered medicinal plant species, said composition comprising modified Murashige and Skoog (MS) basal culture medium, varied concentrations of plant hormones, and other additives, said compositions are;

a. first medium composition useful for initiating fast germination of said plant species with percentage germination ranging between 65 to 98%, and duration period to achieve the same ranging between 7 to 53 days, and also useful for initiating fast release of epicotyl, coleoptile and radicle in percentage seeds ranging between 78 to 100, with duration period for synchronization of the same ranging between 11 to 18 days, said composition consisting of MS basal culture medium, Gibberellic acid (GA_3) ranging between 10 to 100 mg/L,

b. second medium composition useful for initiating fast release of first foliage leaf from epicotyl dormant explants, with percentage secondary explants producing leaves ranging between 65 to 86%, mean number of leaves per secondary explant ranging between 2.7 to 4, and mean leaf length ranging between 3 to 6 cms, said composition consisting of MS basal culture medium, 6- benzyl-aminopurine (BAP) ranging between 3 to 6 mg/L, and Naphthalene acetic acid (NAA) ranging between 0.5 to 1.0 mg/L, and

c. third medium composition useful for initiating fast release of axillary buds from epicotyl dormant explants, with secondary explants producing axillary buds ranging between 70 to 100%, mean number of axillary buds per secondary explant ranging between 6 to 12, and axillary buds producing leaves ranging between 45 to 98%, said composition consisting of MS basal culture medium, 6- benzyl-aminopurine (BAP)

about 2.0 mg/L, Naphthalene acetic acid (NAA) about 1.0 mg/L, and Gibberellic acid (GA3) ranging between 5 to 20 mg/L,

wherein, above-mentioned first medium, second medium, and third medium are used in the same sequence for initiating extraordinarily fast and synchronized *in vitro* induction of germination and release of epicotyl dormancy in plant *Polygonatum cirrhifolium* Royle.

26. Compositions as claimed in claim 25 wherein, plant hormones are selected a from group comprising Gibberellic acid (GA_3), alpha- naphthalene acetic acid (NAA), and 6-benzyl-aminopurine (BAP).

27. Compositions as claimed in claim 25 wherein, Murashige and Skoog's (MS) basal culture medium is preferably consisting of 2.2g/L NH_4NO_3, 2.0g/L KNO_3, 0.44 g/ l $CaCl_2$. $2H_2O$, 0.37 g/L $MgSO_4$. $7H_20$, 0.17g/L KH_2PO_4, 37.25 mg/L Na_2 EDTA, 27.8 mg/L $FeSO_4.7H_20$, 0.83 mg/L KI, 6.2 mg/L H_3 BO_3, 22.3 mg/L $MnSO_4$. $4H_20$, 8.6 mg/L $ZnSO_4$. $7H_20$, 0.25 mg/L Na_2 MoO_4. $2H_20$, 0.025 mg/L $CuSO_4$. $5H_2O$, 0.025 mg/L $CoCl_2.6H_20$, supplemented with 250 mg/L myo-inosital, 0.5mgl/l nicotinic acid, 0.5 mg/L pyridoxine HCl, 3 mg/L thiamine HCl, 2 mg/L glycine, and 30g/L sucrose.

28. Compositions as claimed in claim 25 wherein, first medium composition is used for fast germination, with percentage induction of germination of about 98% is achieved in time duration ranging between 7 to 23 days, said composition consisting of MS basal culture medium and about 50 mg/L Gibberellic acid (GA_3).

29. Compositions as claimed in claim 25 wherein, first medium composition is used for breaking epicotyl dormancy, with about 100% release of epicotyl, coleoptile and radicle, and to achieve the same in time duration ranging between 11 to 14 days, said composition

consisting of MS basal culture medium and Gibberellic acid (GA_3) ranging between 50 to 100 mg/L.

30. Compositions as claimed in claim 25 wherein, second medium composition is used for initiating fast release of first foliage leaf from epicotyl dormant explants, with about 86% secondary explants producing leaves, about 3.4 mean number of leaves per secondary explant, and about 6 cms mean leaf length, said composition consisting of MS basal culture medium, 6- benzyl-aminopurine (BAP) ranging between 3 to 6 mg/L, and Naphthalene acetic acid (NAA) about 1.0 mg/L.

31. Compositions as claimed in claim 25 wherein, third medium composition used for initiating fast release of axillary buds from epicotyl dormant explants, with about 100%, secondary explants producing axillary buds, number of axillary buds per secondary explant ranging between 9 to 12, and axillary buds producing leaves ranging between 77 to 98%, said composition consisting of MS basal culture medium, 6- benzyl-aminopurine (BAP) about 2.0 mg/L, Naphthalene acetic acid (NAA) about 1.0 mg/L, and Gibberellic acid (GA3) ranging between 15 to 20 mg/L.

32. Compositions as claimed in claim 25 wherein, fast *in vitro* multiplication of the said medicinal plant species, with no dormancy period can make this plant commercially available in bulk.

33. Compositions as claimed in claim 25 wherein, said compositions are used for reliable and uniform germination.

34. Compositions as claimed in claim 25 wherein, said compositions are used for initiating faster germination, with only 81 days *in vitro,* to achieve the same.

35. Compositions as claimed in claim 25 wherein, said compositions is the first step towards release of the epicotyl dormancy.

36. Compositions as claimed in claim 25 wherein, said compositions can be used for all possible genotypes of said plant species to be grown *in vitro*.

37. Compositions as claimed in claim 25 wherein, said compositions can be used growing said plant species *in vitro,* in all geographical regions and all seasons.

38. Compositions as claimed in claim 25 wherein, said compositions are used for uniform and high germination rates.

39. Compositions as claimed in claim 25 wherein, said fast *in vitro* multiplication of the said medicinal plant species, is used for conservation of this endangered plant species.

40. Compositions as claimed in claim 25 wherein, said fast *in vitro* multiplication of the said plant species for wider utilization of its medicinal properties.

41. A method for extraordinarily fast and synchronized *in vitro* induction of germination in *Polygonatum cirrhifolium* Royle, said method comprising:

a. obtaining the sterilized seeds from plant *Polygonatum cirrhifolium* Royle,

b. placing the sterilized seeds in sterile disposable plastic Petri plates (10 x 2 cm) containing semi-solid culture medium with 7.5% agar,

c. incubating the parafilm-sealed petri dishes at temperature ranging between 20 to 24^0C and relative humidity (RH) ranging between 50 to 60%,

d. recording the emergence of hypocotyl in the said seeds at an interval of three days,

e. adjusting pH of the medium to 5.8 with 1N NaOH or 1N HCl,

f. sterilizing the medium for 20 minutes at 121°C and 15 lb. psi pressure,

g. dispensing the medium into petri dishes as 30 ml aliquots,

h. incorporating GA_3 into the medium after filter sterilization using 0.22 μm pore size filter to cooled autoclaved medium,

i. transferring said seeds with emerged hypocotyl under aseptic conditions using Laminar Air Flow, to first medium culture consisting of MS basal culture medium, and Gibberellic acid (GA_3) ranging between 10 to 100 mg/L ,

j. incubating the one set of said first medium culture at 30°C under continuos dark conditions (table 1),

k. recording the germination percentage and duration of germination of the said seeds under the first medium culture,

l. incubating the second set of said first culture under diurnal temperature regime of 30/20°C, continuos dark conditions (table 2),

m. recording the germination percentage and duration of germination of the said seeds under second set of first medium culture,

n. transferring the said germinating seeds to third set of first medium culture at 20°C under 16 hours photoperiod in a growth chamber (table 3),

o. recording differentiation of epicotyl with emergent coleoptile and radicle as germination synchronization in third set of first culture medium on daily basis.

42. A method as claimed in claim 41 wherein, first medium composition useful for initiating fast germination of said plant species with percentage germination ranging between 65 to 98%, and duration period to achieve the same ranging between 7 to 53 days, and also useful for initiating fast release of epicotyl, coleoptile and radicle in percentage seeds ranging

between 78 to 100, with duration period for synchronization of the same ranging between 11 to 18 days.

43. A method as claimed in claim 41 wherein, first medium composition is used for fast germination, with percentage induction of germination of about 98% is achieved in time duration ranging between 7 to 23 days, said composition consisting of MS basal culture medium and about 50 mg/L Gibberellic acid (GA_3).

44. A method as claimed in claim 41 wherein, first medium composition is used for breaking epicotyl dormancy, with about 100% release of epicotyl, coleoptile and radicle, and to achieve the same in time duration ranging between 11 to 14 days, said composition consisting of MS basal culture medium and gibberellic acid (GA_3) ranging between 50 to 100 mg/L.

45. A method as claimed in claim 41 wherein, said compositions are used for initiating faster germination, with only 81 days *in vitro,* to achieve the same.

46. A method as claimed in claim 41 wherein, plant hormones are selected a from group comprising gibberellic acid (GA_3), alpha- naphthalene acetic acid (NAA), and 6-benzyl-aminopurine (BAP).

47. A method as claimed in claim 41 wherein, Murashige and Skoog's (MS) basal culture medium is consisting of 2.2g/L NH_4NO_3, 2.0g/L KNO_3, 0.44 g/ l $CaCl_2$. $2H_2O$, 0.37 g/L $MgSO_4$. $7H_20$, 0.17g/L KH_2PO_4, 37.25 mg/L Na_2 EDTA, 27.8 mg/L $FeSO_4.7H_20$, 0.83 mg/L KI, 6.2 mg/L H_3BO_3, 22.3 mg/L $MnSO_4$. $4H_20$, 8.6 mg/L $ZnSO_4$. $7H_20$, 0.25 mg/L Na_2 MoO_4. $2H_20$, 0.025 mg/L $CuSO_4$. $5H_2O$, 0.025 mg/L $CoCl_2.6H_20$, supplemented with 250 mg/L myo-inosital, 0.5mgl/l nicotinic acid, 0.5 mg/L pyridoxine HCl, 3 mg/L thiamine HCl, 2 mg/L glycine, and 30g/L sucrose.

48. A method as claimed in claim 41 wherein, said seeds are washed thoroughly for 2 hours under tap water with 1-2 drops of Tween-20 and rinsed with distilled water.

49. A method as claimed in claim 41 wherein, rinsed seeds are sterilized with 0.1% mercuric chloride ($HgCl_2$).

50. A method as claimed in claim 41 wherein, fast *in vitro* multiplication of the said medicinal plant species, with no dormancy period can make this plant commercially available in bulk.

51. A method as claimed in claim 41 wherein, said compositions are used for reliable and uniform germination.

52. A method as claimed in claim 41 wherein, said compositions can be used for all possible genotypes of said plant species to be grown *in vitro*.

53. A method as claimed in claim 41 wherein, said compositions can be used growing said plant species *in vitro,* in all geographical regions and all seasons.

54. A method as claimed in claim 41 wherein, said compositions are used for uniform and high germination rates.

55. A method as claimed in claim 41 wherein, said fast *in vitro* multiplication of the said medicinal plant species, is used for conservation of this endangered plant species.

56. A method as claimed in claim 41 wherein, said fast *in vitro* multiplication of the said plant species for wider utilization of its medicinal properties.

57. A method for extraordinarily faster release of epicotyl dormancy in *Polygonatum cirrhifolium* Royle using second medium composition consisting of MS basal culture medium, 6- benzyl-aminopurine (BAP) ranging between 3 to 6 mg/L, and Naphthalene acetic acid (NAA) ranging between 0.5 to 1.0 mg/L, said method comprising:

a. incubating the parafilm-sealed petri dishes at temperature ranging between 20 to 24^{0}C and relative humidity (RH) ranging between 50 to 60%,

b. transferring secondary explants consisting of epicotyl with emergent coleoptile and radicle obtained, from said germinating seeds, under aseptic conditions, into the said dishes, using Laminar Air Flow,

c. adjusting pH of the medium to 5.8 with 1N NaOH or 1N HCl,

d. sterilizing the medium for 20 minutes at 121°C and 15 lb. psi pressure,

e. adding BAP and NAA to the above medium to form second medium culture, said composition consisting of MS basal culture medium, 6- benzyl-aminopurine (BAP) ranging between 3 to 6 mg/L, and Naphthalene acetic acid (NAA) ranging between 0.5 to 1.0 mg/L (table 4),

f. dispensing the medium into petri dishes as 30 ml aliquots,

g. incubating the said culture at 20°C under 16 hr photoperiod with light intensity of 2000 lux provided by cool, white fluorescent tubes of 40 watts, and

h. recording the optimal response in release of first foliage leaf from the explant.

58. A method as claimed in claim 57 wherein, second medium composition useful for initiating fast release of first foliage leaf from epicotyl dormant said explants, with percentage secondary explants producing leaves ranging between 65 to 86%, mean number of leaves per secondary explant ranging between 2.7 to 4, and mean leaf length ranging between 3 to 6 cms.

59. A method as claimed in claim 57 wherein, second medium composition is used for initiating fast release of first foliage leaf from said explants, with about 86% secondary explants producing leaves, about 3.4 mean number of leaves per secondary explant, and about 6 cms mean leaf length, said composition consisting of MS basal culture medium, 6-

benzyl-aminopurine (BAP) ranging between 3 to 6 mg/L, and Naphthalene acetic acid (NAA) about 1.0 mg/L.

60. A method as claimed in claim 57 wherein, plant hormones are selected a from group comprising gibberellic acid (GA_3), alpha- naphthalene acetic acid (NAA), and 6-benzyl-aminopurine (BAP).

61. A method as claimed in claim 57 wherein, Murashige and Skoog's (MS) basal culture medium is consisting of 2.2g/L NH_4NO_3, 2.0g/L KNO_3, 0.44 g/ l $CaCl_2$. $2H_2O$, 0.37 g/L $MgSO_4$. $7H_20$, 0.17g/L KH_2PO_4, 37.25 mg/L Na_2 EDTA, 27.8 mg/L $FeSO_4.7H_20$, 0.83 mg/L KI, 6.2 mg/L H_3 BO_3, 22.3 mg/L $MnSO_4$. $4H_20$, 8.6 mg/L $ZnSO_4$. $7H_20$, 0.25 mg/L Na_2 MoO_4. $2H_20$, 0.025 mg/L $CuSO_4$. $5H_2O$, 0.025 mg/L $CoCl_2.6H_20$, supplemented with 250 mg/L myo-inosital, 0.5mgl/l nicotinic acid, 0.5 mg/L pyridoxine HCl, 3 mg/L thiamine HCl, 2 mg/L glycine, and 30g/L sucrose.

62. A method as claimed in claim 57 wherein, said seeds are washed thoroughly for 2 hours under tap water with 1-2 drops of Tween-20™ and rinsed with distilled water.

63. A method as claimed in claim 57 wherein, rinsed seeds are sterilized with 0.1% mercuric chloride ($HgCl_2$).

64. A method as claimed in claim 57 wherein, fast *in vitro* multiplication of the said medicinal plant species, with no dormancy period can make this plant commercially available in bulk.

65. A method as claimed in claim 57 wherein, said compositions are used for reliable and uniform germination.

66. A method as claimed in claim 57 wherein, said compositions can be used for all possible genotypes of said plant species to be grown *in vitro*.

67. A method as claimed in claim 57 wherein, said compositions can be used growing said plant species *in vitro,* in all geographical regions and all seasons.

68. A method as claimed in claim 57 wherein, said compositions are used for uniform and high germination rates.

69. A method as claimed in claim 57 wherein, said fast *in vitro* multiplication of the said medicinal plant species, is used for conservation of this endangered plant species.

70. A method as claimed in claim 57 wherein, said fast *in vitro* multiplication of the said plant species for wider utilization of its medicinal properties.

71. A method for extraordinarily faster release of epicotyl dormancy from differentiated de novo axillary bud and release of foliage leaves in *Polygonatum cirrhifolium* Royle, using third medium composition consisting of MS basal culture medium, 6- benzyl-aminopurine (BAP) about 2.0 mg/L, Naphthalene acetic acid (NAA) about 1.0 mg/L, and Gibberellic acid (GA3) ranging between 5 to 20 mg/L, said method comprising:

a. incorporating GA_3 into the fresh MS basal culture medium with NAA and BAP, after filter sterilization using 0.22 μm pore size filter to cooled autoclaved medium, to form third medium culture, said composition consisting,

b. transferring fresh secondary explants consisting of epicotyl with emergent coleoptile and radicle obtained from said germinating seeds explant under aseptic conditions using Laminar Air Flow, to third culture medium,

c. incubating the said culture at about 20°C under duration ranging between 10 to 6 hr photoperiod, with light intensity of about 2000 lux provided by cool,

d. maintaining the incubated cultures at 50-60% RH,

e. subculturing the said culture after every four weeks on the third medium formulations as mentioned above,

f. recording the de novo axillary bud differentiation and release of foliage leaves from them, and

g. compiling the final data after 16 weeks of culture on the basis of periodical observations.

72. A method as claimed in claim 71 wherein, third medium composition useful for initiating fast release of axillary buds from epicotyl dormant said explants, with secondary explants producing axillary buds ranging between 70 to 100%, mean number of axillary buds per secondary explant ranging between 6 to 12, and axillary buds producing leaves ranging between 45 to 98%.

73. A method as claimed in claim 71 wherein, third medium composition used for initiating fast release of axillary buds from said explants, with about 100%, secondary explants producing axillary buds, number of axillary buds per secondary explant ranging between 9 to 12, and axillary buds producing leaves ranging between 77 to 98%, said composition consisting of MS basal culture medium, 6- benzyl-aminopurine (BAP) about 2.0 mg/L, Naphthalene acetic acid (NAA) about 1.0 mg/L, and Gibberellic acid (GA3) ranging between 15 to 20 mg/L.

74. A method as claimed in claim 71 wherein, plant hormones are selected a from group comprising gibberellic acid (GA_3), alpha- naphthalene acetic acid (NAA), and 6-benzyl-aminopurine (BAP).

75. A method as claimed in claim 71 wherein, Murashige and Skoog's (MS) basal culture medium is consisting of 2.2g/L NH_4NO_3, 2.0g/L KNO_3, 0.44 g/ l $CaCl_2$. $2H_2O$, 0.37 g/L

$MgSO_4$. $7H_20$, 0.17g/L KH_2PO_4, 37.25 mg/L Na_2 EDTA, 27.8 mg/L $FeSO_4.7H_20$, 0.83 mg/L KI, 6.2 mg/L H_3 BO_3, 22.3 mg/L $MnSO_4$. $4H_20$, 8.6 mg/L $ZnSO_4$. $7H_20$, 0.25 mg/L Na_2 MoO_4. $2H_20$, 0.025 mg/L $CuSO_4$. $5H_2O$, 0.025 mg/L $CoCl_2.6H_20$, supplemented with 250 mg/L myo-inosital, 0.5mgl/l nicotinic acid, 0.5 mg/L pyridoxine HCl, 3 mg/L thiamine HCl, 2 mg/L glycine, and 30g/L sucrose.

76. A method as claimed in claim 71 wherein, said seeds are washed thoroughly for 2 hours under tap water with 1-2 drops of Tween-20 and rinsed with distilled water.

77. A method as claimed in claim 71 wherein, rinsed seeds are sterilized with 0.1% mercuric chloride ($HgCl_2$).

78. A method as claimed in claim 71 wherein, fast *in vitro* multiplication of the said medicinal plant species, with no dormancy period can make this plant commercially available in bulk.

79. A method as claimed in claim 71 wherein, said compositions are used for reliable and uniform germination.

80. A method as claimed in claim 71 wherein, said compositions can be used for all possible genotypes of said plant species to be grown *in vitro.*

81. A method as claimed in claim 71 wherein, said compositions can be used growing said plant species *in vitro,* in all geographical regions and all seasons.

82. A method as claimed in claim 71 wherein, said compositions are used for uniform and high germination rates.

83. A method as claimed in claim 71 wherein, said fast *in vitro* multiplication of the said medicinal plant species, is used for conservation of this endangered plant species.

84. A method as claimed in claim 71 wherein, said fast *in vitro* multiplication of the said plant species for wider utilization of its medicinal properties.